

**Table 2B-1. Zone Diameter and MIC Breakpoints for *Pseudomonas aeruginosa***

<p><b>Testing Conditions</b></p> <p><b>Medium:</b> Disk diffusion: MHA  Broth dilution: CAMHB; iron-depleted CAMHB for cefiderocol (see Appendix I)<sup>1</sup>  Agar dilution: MHA</p> <p><b>Inoculum:</b> Broth culture method or colony suspension, equivalent to a 0.5 McFarland standard</p> <p><b>Incubation:</b> 35°C ± 2°C; ambient air  Disk diffusion: 16-18 hours  Dilution methods: 16-20 hours</p>	<p><b>Routine QC Recommendations</b> (see Tables 4A-1 and 5A-1 for acceptable QC ranges)</p> <p><i>Pseudomonas aeruginosa</i> ATCC<sup>®a</sup> 27853</p> <p>Refer to Tables 4A-2 and 5A-2 to select strains for routine QC of β-lactam combination agents.</p> <p>When a commercial test system is used for susceptibility testing, refer to the manufacturer’s instructions for QC test recommendations and QC ranges.</p>
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**General Comments**

- (1) For disk diffusion, test a maximum of 12 disks on a 150-mm plate and no more than 6 disks on a 100-mm plate; disks should be placed no less than 24 mm apart, center to center (see M02,<sup>2</sup> Subchapter 3.6). Each zone diameter should be clearly measurable; overlapping zones prevent accurate measurement. Measure the diameter of the zones of complete inhibition (as judged by the unaided eye), including the diameter of the disk (see the *M02 Disk Diffusion Reading Guide*<sup>3</sup>). Hold the Petri plate a few inches above a black background illuminated with reflected light. The zone margin should be considered the area showing no obvious, visible growth that can be detected with the unaided eye. Ignore faint growth of tiny colonies that can be detected only with a magnifying lens at the edge of the zone of inhibited growth.
- (2) The susceptibility of *P. aeruginosa* isolated from patients with cystic fibrosis can be reliably determined by disk diffusion or dilution methods but may need extended incubation for up to 24 hours before reporting as susceptible.
- (3) *P. aeruginosa* may develop resistance during prolonged therapy with all antimicrobial agents. Therefore, isolates that are initially susceptible may become resistant within 3 to 4 days after initiation of therapy. Testing of repeat isolates may be warranted.
- (4) The dosage regimens shown in the comments column below are those necessary to achieve plasma drug exposures (in adults with normal renal and hepatic functions) on which breakpoints were derived. When implementing new breakpoints, it is strongly recommended that laboratories share this information with infectious diseases practitioners, pharmacists, pharmacy and therapeutics committees, infection prevention committees, and the antimicrobial stewardship team.
- (5) Intermediate ranges denoted with a ^ for the applicable antimicrobial agents in the drug groups in Tables 2 are based on the known ability of these agents to concentrate in the urine.

**NOTE:** Information in boldface type is new or modified since the previous edition.

**Table 2B-1. *Pseudomonas aeruginosa* (Continued)**

Test/Report Group	Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Interpretive Categories and MIC Breakpoints, µg/mL			Comments
			S	I	R	S	I	R	
<b>PENICILLINS</b>									
O	Piperacillin	100 µg	≥21	15-20 <sup>^</sup>	≤14	≤16	32-64 <sup>^</sup>	≥128	(6) Breakpoints for piperacillin (alone or with tazobactam) are based on a piperacillin dosage regimen of at least 3 g administered every 6 h.
<b>β-LACTAM COMBINATION AGENTS</b>									
A	Piperacillin-tazobactam	100/10 µg	≥21	15-20 <sup>^</sup>	≤14	≤16/4	32/4-64/4 <sup>^</sup>	≥128/4	(7) Breakpoints for piperacillin (alone or with tazobactam) are based on a piperacillin dosage regimen of at least 3 g administered every 6 h.
B	Ceftazidime-avibactam	30/20 µg	≥21	-	≤20	≤8/4	-	≥16/4	(8) Breakpoints are based on a dosage regimen of 2.5 g administered every 8 h over 2 h.
B	Ceftolozane-tazobactam	30/10 µg	≥21	17-20 <sup>^</sup>	≤16	≤4/4	8/4 <sup>^</sup>	≥16/4	(9) Breakpoints are based on a dosage regimen of 1.5 g administered every 8 h.
B	Imipenem-relebactam	10/25 µg	≥23	20-22 <sup>^</sup>	≤19	≤2/4	4/4 <sup>^</sup>	≥8/4	(10) Breakpoints are based on a dosage regimen of 1.25 g administered every 6 h.  (11) Organisms that test susceptible to imipenem are also considered susceptible to imipenem-relebactam. However, organisms that test susceptible to imipenem-relebactam cannot be assumed to be susceptible to imipenem.
O	Ticarcillin-clavulanate	75/10 µg	≥24	16-23 <sup>^</sup>	≤15	≤16/2	32/2-64/2 <sup>^</sup>	≥128/2	(12) Breakpoints for ticarcillin (alone or with clavulanate) are based on a ticarcillin dosage regimen of at least 3 g administered every 6 h.
<b>CEPHEMS (PARENTERAL) (Including cephalosporins I, II, III, and IV. Please refer to Glossary I.)</b>									
A	Ceftazidime	30 µg	≥18	15-17 <sup>^</sup>	≤14	≤8	16 <sup>^</sup>	≥32	(13) Breakpoints are based on a dosage regimen of 1 g administered every 6 h or 2 g administered every 8 h.
B	Cefepime	30 µg	≥18	15-17 <sup>^</sup>	≤14	≤8	16 <sup>^</sup>	≥32	(14) Breakpoints are based on a dosage regimen of 1 g administered every 8 h or 2 g administered every 12 h.
Inv.	Cefiderocol	30 µg	≥18	13-17 <sup>^</sup>	≤12	≤4	8 <sup>^</sup>	≥16	(15) Breakpoints are based on a dosage regimen of 2 g every 8 h administered over 3 h.
<b>MONOBACTAMS</b>									
B	Aztreonam	30 µg	≥22	16-21 <sup>^</sup>	≤15	≤8	16 <sup>^</sup>	≥32	(16) Breakpoints are based on a dosage regimen of 1 g administered every 6 h or 2 g administered every 8 h.

Table 2B-1. *Pseudomonas aeruginosa* (Continued)

Test/Report Group	Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Interpretive Categories and MIC Breakpoints, µg/mL			Comments
			S	I	R	S	I	R	
<b>CARBAPENEMS</b>									
B	Doripenem	10 µg	≥19	16-18 <sup>^</sup>	≤15	≤2	4 <sup>^</sup>	≥8	(17) Breakpoints for doripenem are based on a dosage regimen of 500 mg administered every 8 h.
	Imipenem	10 µg	≥19	16-18 <sup>^</sup>	≤15	≤2	4 <sup>^</sup>	≥8	(18) Breakpoints for imipenem are based on a dosage regimen of 1 g administered every 8 h or 500 mg administered every 6 h.  See comment (11).
	Meropenem	10 µg	≥19	16-18 <sup>^</sup>	≤15	≤2	4 <sup>^</sup>	≥8	(19) Breakpoints for meropenem are based on a dosage regimen of 1 g administered every 8 h.
<b>LIPOPEPTIDES</b>									
(20) <b>WARNING:</b> Clinical and PK/PD data demonstrate colistin and polymyxin B have limited clinical efficacy, even if an intermediate result is obtained. Alternative agents are strongly preferred. Colistin and polymyxin B should be used in combination with one or more active antimicrobial agents. Consultation with an infectious diseases specialist is recommended.									
O	Colistin or polymyxin B	-	-	-	-	-	≤2	≥4	(21) Colistin (methanesulfonate) should be given with a loading dose and maximum renally adjusted doses (see International Consensus Guidelines <sup>4</sup> ).  (22) Polymyxin B should be given with a loading dose and maximum recommended doses (see International Consensus Guidelines <sup>4</sup> ).  (23) When colistin or polymyxin B is given systemically, neither is likely to be effective for pneumonia.  (24) For colistin, broth microdilution, CBDE, and CAT MIC methods are acceptable. For polymyxin B, broth microdilution is the only approved method. Disk diffusion and gradient diffusion methods should not be performed (see Table 3D).

**Table 2B-1. *Pseudomonas aeruginosa* (Continued)**

Test/Report Group	Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Interpretive Categories and MIC Breakpoints, µg/mL			Comments
			S	I	R	S	I	R	
<b>AMINOGLYCOSIDES</b>									
A	Gentamicin	10 µg	≥ 15	13-14 <sup>^</sup>	≤ 12	≤ 4	8 <sup>^</sup>	≥ 16	
A	Tobramycin	10 µg	≥ 15	13-14 <sup>^</sup>	≤ 12	≤ 4	8 <sup>^</sup>	≥ 16	
B	Amikacin	30 µg	≥ 17	15-16 <sup>^</sup>	≤ 14	≤ 16	32 <sup>^</sup>	≥ 64	
O	Netilmicin	30 µg	≥ 15	13-14 <sup>^</sup>	≤ 12	≤ 8	16 <sup>^</sup>	≥ 32	
<b>FLUOROQUINOLONES</b>									
B	Ciprofloxacin	5 µg	≥ 25	19-24 <sup>^</sup>	≤ 18	≤ 0.5	1 <sup>^</sup>	≥ 2	(25) Breakpoints are based on a dosage regimen of 400 mg IV administered every 8 h.
B	Levofloxacin	5 µg	≥ 22	15-21 <sup>^</sup>	≤ 14	≤ 1	2 <sup>^</sup>	≥ 4	(26) Breakpoints are based on a dosage regimen of 750 mg administered every 24 h.
O	Lomefloxacin	10 µg	≥ 22	19-21 <sup>^</sup>	≤ 18	≤ 2	4 <sup>^</sup>	≥ 8	(27) For testing and reporting of urinary tract isolates only.
O	Norfloxacin	10 µg	≥ 17	13-16	≤ 12	≤ 4	8	≥ 16	See comment (27).
O	Ofloxacin	5 µg	≥ 16	13-15 <sup>^</sup>	≤ 12	≤ 2	4 <sup>^</sup>	≥ 8	
O	Gatifloxacin	5 µg	≥ 18	15-17 <sup>^</sup>	≤ 14	≤ 2	4 <sup>^</sup>	≥ 8	

Abbreviations: ATCC®, American Type Culture Collection; CAMHB, cation-adjusted Mueller-Hinton broth; CAT, colistin agar test; CBDE, colistin broth disk elution; I, intermediate; IV, intravenous; MHA, Mueller-Hinton agar; MIC, minimal inhibitory concentration; PK/PD, pharmacokinetic/pharmacodynamic; QC, quality control; R, resistant; S, susceptible.

Symbol: <sup>^</sup>, designation for agents that have the potential to concentrate in the urine.

**Footnote**

- a. ATCC® is a registered trademark of the American Type Culture Collection.

**References for Table 2B-1**

- 1 Hackel MA, Tsuji M, Yamano Y, Echols R, Karlowsky JA, Sahm DF. Reproducibility of broth microdilution MICs for the novel siderophore cephalosporin, cefiderocol, determined using iron-depleted cation-adjusted Mueller-Hinton broth. *Diagn Microbiol Infect Dis*. 2019;94(4):321-325.
- 2 CLSI. *Performance Standards for Antimicrobial Disk Susceptibility Tests*. 13th ed. CLSI standard M02. Clinical and Laboratory Standards Institute; 2018.
- 3 CLSI. *M02 Disk Diffusion Reading Guide*. 1st ed. CLSI quick guide M02QG. Clinical and Laboratory Standards Institute; 2018.
- 4 Tsuji BT, Pogue JM, Zavascki AP, et al. International consensus guidelines for the optimal use of the polymyxins: endorsed by the American College of Clinical Pharmacy (ACCP), European Society of Clinical Microbiology and Infectious Diseases (ESCMID), Infectious Diseases Society of America (IDSA), International Society for Anti-Infective Pharmacology (ISAP), Society of Critical Care Medicine (SCCM), and Society of Infectious Diseases Pharmacists (SIDP). *Pharmacotherapy*. 2019;39(1):10-39.